



# Comparison of homeopathic globules prepared from high and ultra-high dilutions of various starting materials by ultraviolet light spectroscopy



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## ABSTRACT

**Objective:** Homeopathic globules are commonly used in clinical practice, while research focuses on liquid potencies. Sequential dilution and succussion in their production process has been proposed to change the physico-chemical properties of the solvent(s). It has been reported that aqueous potencies of various starting materials showed significant differences in ultraviolet light transmission compared to controls and between different dilution levels. The aim of the present study was to repeat and expand these experiments to homeopathic globules.

**Methods:** Globules were specially produced for this study by Spagyros AG (Gümligen, Switzerland) from 6 starting materials (*Aconitum napellus*, *Atropa belladonna*, phosphorus, sulfur, *Apis mellifica*, quartz) and for 6 dilution levels (6x, 12x, 30c, 200c, 200CF (centesimal discontinuous fluxion), 10,000CF). Native globules and globules impregnated with solvents were used as controls. Globules were dissolved in ultrapure water, and absorbance in the ultraviolet range was measured. The average absorbance from 200 to 340 nm was calculated and corrected for differences between measurement days and instrumental drift.

**Results:** Statistically significant differences were found for *A. napellus*, sulfur, and *A. mellifica* when normalized average absorbance of the various dilution levels from the same starting material (including control and solvent control globules) was compared. Additionally, absorbance within dilution levels was compared among the various starting materials. Statistically significant differences were found among 30c, 200c and 200CF dilutions.

**Conclusion:** This study has expanded previous findings from aqueous potencies to globules and may indicate that characteristics of aqueous high dilutions may be preserved and detectable in dissolved globules.

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## 1. Introduction

Highly diluted remedies are applied in homeopathy and anthroposophically extended medicine. Several modes of action and models have been discussed on how the production of these aqueous solutions by sequential dilution and succussion, also termed potentization, might feature their structural properties.<sup>1</sup> It has been proposed that a network of hydrogen bonds develops around non-polar solutes that remains with successive dilution, even when the molecules of the starting material have disappeared,<sup>2,3</sup> thus giving

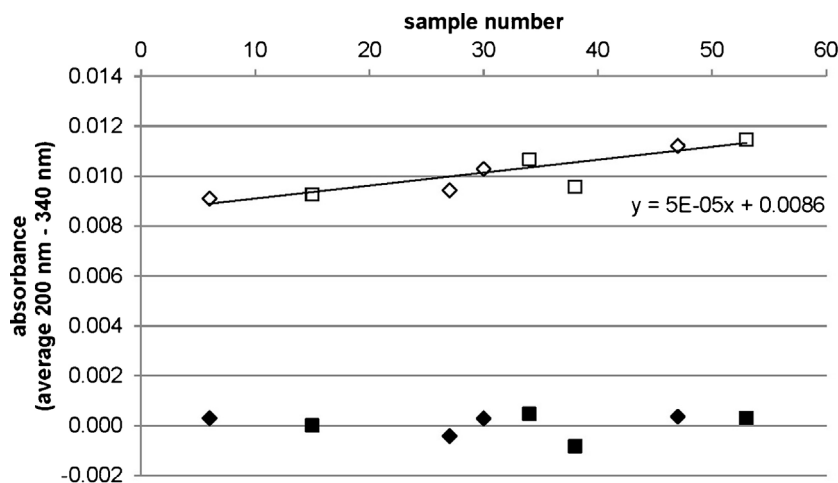
the water a more organized state.<sup>4</sup> Additionally, nanobubbles have been discussed to contribute to these supramolecular structures,<sup>5</sup> which are assumed to disappear upon heating.<sup>4</sup>

Differences in the structure of the water in these homeopathic preparations may be reflected in small but measurable changes in physico-chemical properties. Accordingly, methods such as ultraviolet (UV) spectroscopy,<sup>2,6–12</sup> nuclear magnetic resonance spectroscopy,<sup>4,13–17</sup> calorimetry<sup>18</sup> or thermoluminescence<sup>19,20</sup> have been employed to investigate possible differences between homeopathic preparations and respective controls.

It has been found that dilution and succussion can lead to the introduction of contaminants, e.g., trace elements such as Si, Li, Na, Mg.<sup>9,21</sup> This can be avoided when dilutions are prepared very carefully (e.g., by washing all flasks and pipettes with the same solution used to prepare the dilutions) or under clean room conditions.<sup>16,22</sup>

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**Fig. 1.** Obtaining normalized absorbance by correcting for instrumental drift.

A typical example from one measurement day is shown. Diamonds represent native globules, and squares represent solvent control globules. Open symbols show the average absorbance from 200 nm to 340 nm. Samples were measured in a randomized order, and plotting absorbance versus sample number revealed an instrumental drift over time. The intercept of the linear regression (measure for the differences between measurement days) and the slope (measure for the instrumental drift) were used to correct the values, yielding the normalized absorbance (filled symbols). The same equation was then used to normalize the absorbance of the various dilution levels measured on the same day (not shown). A normalized absorbance can thus have positive or negative values.

This is crucial for experiments testing for differences between high dilutions and controls. Our group reported that high and ultra-high dilutions of various starting materials showed significant differences in UV light transmission to controls and between different dilution levels, all the more so when these dilutions were produced under controlled conditions.<sup>10–12</sup>

In clinical practice, globules of potentized starting materials are commonly applied besides aqueous dilutions. In a preliminary study, we found differences in UV absorbance between verum and placebo globules of *Aconitum napellus* 30c or calcium carbonate/querqus e cortice 6x dissolved in water.<sup>23</sup> These globules had been produced for clinical trials and not for laboratory experiments. It was unclear whether the differences in UV absorbance originated from specific characteristics of the starting materials, from differences in the production of verum and placebo globules, and/or other unknown interference factors.

Therefore, the aim of the present study was to repeat and expand our previous experiments with globules produced under controlled conditions accurate for comparison of UV absorbance, i.e., sucrose globules and ethanol from the same batch were used to minimize the introduction of possible artifacts.

## 2. Methods

### 2.1. Globules

The globules were specially produced for this study by Spagyros AG (Gümligen, Switzerland) and differed only in the starting materials of the potentized dilutions. Sucrose globules and ethanol from the same batch were used to minimize the introduction of possible artifacts. The following starting materials were investigated: *A. napellus*, *Atropa belladonna*, phosphorus, sulfur, *Apis mellifica*, and quartz (2 plants, 2 non-metal elements, 1 animal, and 1 mineral).

Dilution was performed in a 43% ethanol/57% water mixture until the last step, for which 73% ethanol/27% water was used. Manual succussion was performed vertically with 30 strokes in each step.

200CF and 10,000CF dilutions were produced by discontinuous fluxion in a machine with water as dilution medium. In this single glass technique, the potentisation vessel is alternately filled with the dilution medium and emptied again in each step, while

a defined amount of liquid remains in the vessel. The operating mode of the machine has been described.<sup>24</sup> The final three steps were manually diluted in 30% ethanol/70% water, 43% ethanol/57% water and 73% ethanol/27% water and succussed with 30 strokes.

Two kinds of controls were generated in the experiments: native globules and globules impregnated with a succussed 73% ethanol/27% water mixture (solvent control globules).

The globules were produced in July 2012 and the measurements were carried out from September 2012 to March 2013. Vials were coded to display the starting material but the dilution level and the controls were blinded. Coding was unblinded only after completion of all measurements and preliminary description of the data by box plots. The vials were stored in aluminum boxes, each box containing the same dilution level of the 6 starting materials.

### 2.2. UV absorbance measurements

Globules were gently dissolved in ultrapure water (arium® pro VF, Sartorius Stedim AG, Goettingen, Germany) at 10 mg/ml in Fiolax® test tubes. Samples were prepared in quadruplicates, 19–22 h prior to the measurements to allow complete dissolution, wrapped individually in aluminum foil and stored in the dark at room temperature. Absorbance of the samples in the UV range (from 190 to 340 nm) was measured in a randomized order with a Shimadzu UV-1800 double beam spectrophotometer (Reinach, Switzerland) equipped with an auto sampler CETAC ASX-260 (Omaha, USA; as described previously in Ref.<sup>10</sup>). This wavelength range has previously proven to be better suited to detect differences than visible light or near infrared light.<sup>25</sup> The spectrophotometer had been switched on 2 h prior to the measurements for sufficient warming up. Globules of each starting material and corresponding controls were freshly dissolved and measured on 5 independent days.

### 2.3. Data analysis

Due to the high absorbance of sucrose below 200 nm, the main component of the globules used, only absorbance values at 200 nm and higher were included in the analysis and one value for each nm was recorded. For each sample the mean of the absorbance values from 200 nm to 340 nm was calculated. Fig. 1 shows how

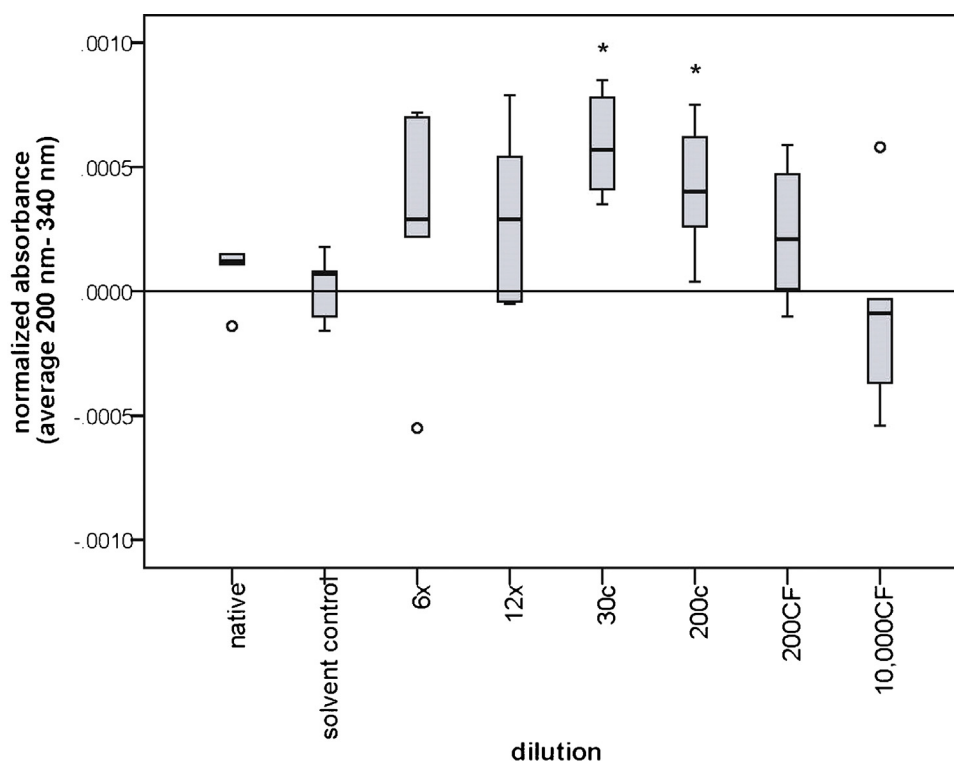
**Table 1**  
Comparison<sup>a</sup> of normalized absorbance<sup>b</sup> of globules of various dilution levels<sup>c</sup> for each starting material.

	Including native and solvent control globules		Including solvent control globules	
	H (7)	<i>p</i>	H (6)	<i>p</i>
<i>Aconitum napellus</i>	14.31	<b>.046</b>	11.68	.070
<i>Atropa belladonna</i>	4.88	.675	3.73	.713
Phosphorus	2.52	.925	1.48	.961
Sulfur	15.24	<b>.033</b>	9.52	.147
<i>Apis mellifica</i>	14.71	<b>.040</b>	13.59	<b>.035</b>
Quartz	4.89	.673	4.12	.660

<sup>a</sup> By Kruskal–Wallis test, statistically significant results ( $p \leq 0.05$ ) are displayed in bold, H = test statistic with degrees of freedom in parentheses.

<sup>b</sup> Average between 200 nm and 340 nm.

<sup>c</sup> 6x, 12x, 30c, 200c, 200CF, 10,000CF, control(s) as indicated ( $n = 5$  for each starting material and dilution level).



**Fig. 2.** Boxplot showing normalized absorbance of control and *Aconitum napellus* globules of various dilution levels.

Globules were dissolved in water, their absorbance was measured and corrected for the daily variations and drift in the spectrophotometer, yielding a normalized average absorbance from 200 nm to 340 nm ( $n = 5$  for each dilution level). Circles represent outliers (that lie more than one and a half box lengths above or below from the upper or lower quartile, respectively). \*Indicate statistically significant differences ( $p \leq 0.05$ , Mann–Whitney–U test) between the respective dilution level and the solvent control globules.

this value was corrected for the differences between measurement days and the instrumental drift during the measurements by linear regression (intercept as measure for the differences between measurement days and slope as measure for the drift), yielding a normalized absorbance. Native and solvent control globules were thereby regarded as references, since they were independent of the starting materials.

Since some data were not normally distributed, non-parametric statistical tests were applied. The normalized average absorbance from 200 to 340 nm was compared between various starting materials within equal dilution levels and between various dilution levels within equal starting materials using a Kruskal–Wallis test. Within equal starting materials, each dilution level was compared to the solvent control globules using a Mann–Whitney–U test.  $p \leq 0.05$  was considered statistically significant. Effect sizes ( $r$ ) were calculated and results were reported according to Ref. <sup>26</sup> no adjustments for multiple comparisons were performed. While such adjustments (e.g., Bonferroni adjustments) decrease the likelihood

for type I errors (mistakenly rejecting the null hypothesis), they increase the likelihood for type II errors, i.e., important differences are deemed non-significant.<sup>27,28</sup>

Statistical analysis was performed with IBM SPSS Statistics 21.0 (Armonk, NY, USA).

### 3. Results

Normalized average absorbances of the various dilution levels and native and solvent control globules from the same starting material were compared by a Kruskal–Wallis test. Statistically significant differences were found for *A. napellus*, sulfur, and *A. mellifica* (Table 1). The example of *A. napellus* is illustrated in Fig. 2.

Native and solvent control globules showed a statistically significant difference from each other ( $p = 0.035$ ,  $r = -0.272$ ). Absorbance of dilution levels were then individually compared to the solvent control globules, which were considered a more accurate control than native globules that had not been impregnated with ethanol

**Table 2**  
Comparison<sup>a</sup> between normalized absorbance<sup>b</sup> of solvent control globules and globules of various starting materials and dilution levels<sup>c</sup>.

		<i>Aconitum napellus</i>	<i>Atropa belladonna</i>	Phosphorus	Sulfur	<i>Apis mellifica</i>	Quartz
Solvent control	Median	.00007	.00003	-.00016	-.00001	-.00038	-.00003
	MAD	.00011	.00016	.00004	.00008	.00027	.00015
6x	Median	.00029	.00012	-.00011	.00002	-.00017	.00005
	MAD	.00041	.00031	.00023	.00023	.00013	.00003
	<i>p</i>	.117	.465	.834	.602	.175	.917
	<i>r</i>	-.496	-.231	-.066	-.165	-.429	-.033
12x	Median	.00029	-.00020	-.00016	-.00014	.00039	-.00011
	MAD	.00033	.00028	.00035	.00007	.00072	.00033
	<i>p</i>	.175	.754	.675	.602	.251	.754
	<i>r</i>	-.429	-.099	-.132	-.165	-.363	-.099
30c	Median	.00057	-.00009	-.00012	-.00011	.00020	-.00006
	MAD	.00021	.00044	.00006	.00007	.00022	.00011
	<i>p</i>	<b>.009</b>	.834	.465	.344	<b>.028</b>	.754
	<i>r</i>	-.826	-.066	-.231	-.299	-.693	-.099
200c	Median	.00040	-.00047	-.00011	-.00022	.00036	.00007
	MAD	.00022	.00039	.00018	.00008	.00017	.00080
	<i>p</i>	<b>.047</b>	.249	.834	.465	<b>.028</b>	.917
	<i>r</i>	-.627	-.364	-.066	-.231	-.693	-.033
200CF	Median	.00021	-.00008	-.00019	.00022	.00012	.00002
	MAD	.00026	.00006	.00005	.00006	.00016	.00006
	<i>p</i>	.209	.530	.754	<b>.009</b>	.076	.917
	<i>r</i>	-.397	-.199	-.099	-.826	-.562	-.033
10,000CF	Median	-.00009	-.00042	-.00020	-.00009	.00032	.00048
	MAD	.00028	.00017	.00011	.00024	.00010	.00013
	<i>p</i>	.465	.249	.675	.917	<b>.009</b>	.117
	<i>r</i>	-.231	-.364	-.132	-.033	-.826	-.496

<sup>a</sup> By Mann–Whitney–U test, median normalized absorbance with median absolute deviation (MAD) is shown, statistically significant results ( $p \leq 0.05$ ) are displayed in bold, *r* = effect size.

<sup>b</sup> Average between 200 nm and 340 nm.

<sup>c</sup>  $n = 5$  for each starting material and dilution level.

**Table 3**  
Comparison<sup>a</sup> of normalized absorbance<sup>b</sup> of globules of various starting materials<sup>c</sup> for each dilution level.

	H (5)	<i>p</i>
Native	13.59	<b>.018</b>
Solvent control	4.93	.424
6x	4.66	.459
12x	3.82	.575
30c	11.36	<b>.045</b>
200c	11.94	<b>.036</b>
200CF	11.95	<b>.036</b>
10,000CF	9.97	.076

<sup>a</sup> By Kruskal–Wallis test, statistically significant results ( $p \leq 0.05$ ) are displayed in bold, H = test statistic with degrees of freedom in parentheses. Pairwise comparisons of the 6 starting materials were not performed due to the high probability of a type I error when doing 15 tests.

<sup>b</sup> Average between 200 nm and 340 nm.

<sup>c</sup> *Aconitum napellus*, *Atropa belladonna*, phosphorus, sulfur, *Apis mellifica*, quartz ( $n = 5$  for each starting material and dilution level).

and water. Statistically significant differences were found between several of the ultra-high dilutions of *A. napellus*, sulfur, and *A. mellifica* and the respective solvent control globules (Table 2).

Finally, absorbance within dilution levels were compared between the various starting materials. Statistically significant differences were found between 30c, 200c and 200CF dilutions and between control globules (Table 3). The example of 200CF globules is illustrated in Fig. 3. No differences were found between 6x, 12x and 10,000CF dilutions and among solvent control globules.

#### 4. Discussion

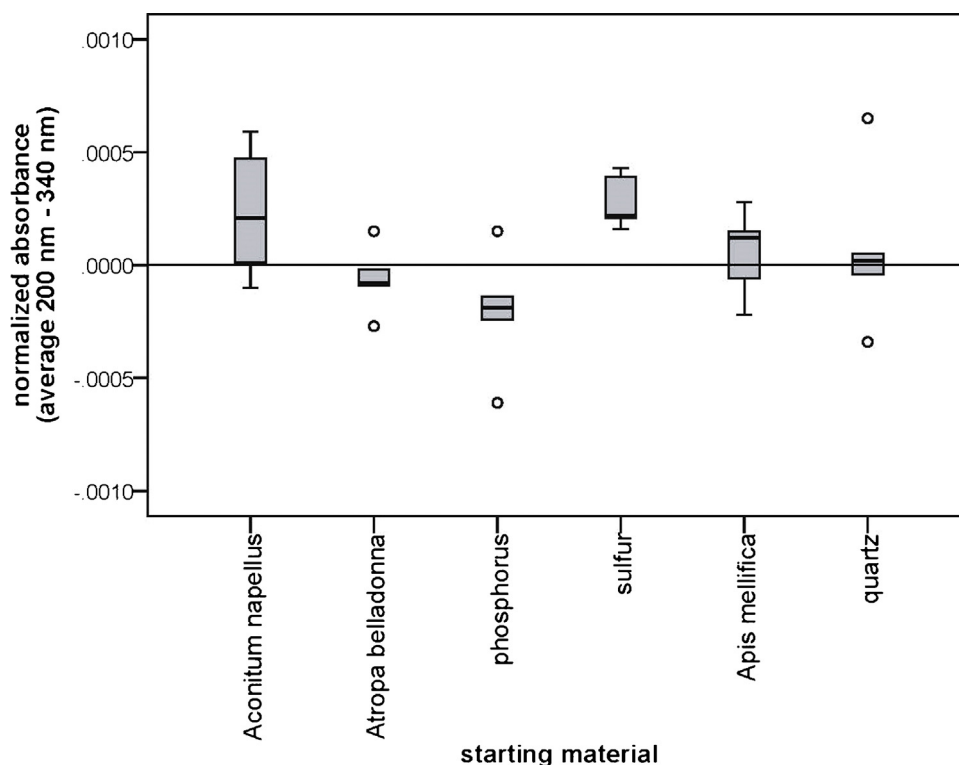
To the best of our knowledge, this is a logical development of our previous study,<sup>23</sup> the first report on differences in UV absorbance between globules prepared from various starting materials and

various dilution levels. Apart from differences between control globules and globules impregnated with defined dilution levels, we found significant differences for 30c, 200c and 200CF from various starting materials, which indicates that the starting material is important in the potentization process, although at these high dilutions a presence of molecules of the starting material is not expected. This is in agreement with Rey, who was able to distinguish between lithium chloride 15c and sodium chloride 15c by thermoluminescence.<sup>19</sup>

Limitations of UV absorbance measurements of high dilutions in general include e.g., the detection limit of the spectrophotometer or differences in absolute absorbance between measurement days, and they have been discussed previously.<sup>12</sup> In the present study, we corrected for the daily variations of the instrument as well as the drift during the measurements. Additionally, the globules were produced under controlled state-of-the-art conditions and in accordance with legal regulations by a pharmaceutical company. The results were generally consistent with former studies of aqueous dilutions, although by freshly weighing in and dissolving the globules for each measurement, an additional preparatory step was introduced, which may have added to the variations between measurements.

Each series of dilutions from each starting material was measured on 5 different days. Thus, we were limited in statistical testing, i.e., pairwise comparisons between starting materials were not performed and Bonferroni corrections were not applied. In future studies, one could focus on fewer starting materials and dilution levels and perform more measurements.

We used high dilutions to impregnate globules, which were then dissolved in water. In a future study it would be worthwhile to not only measure absorbance of the dissolved globules but also of the original high dilutions, i.e., the solutions with which the globules were impregnated, in order to investigate whether comparable dif-



**Fig. 3.** Boxplot showing normalized absorbance of 200CF globules from various starting materials.

Globules were dissolved in water, their absorbance was measured and corrected for the daily variations and drift in the spectrophotometer, yielding a normalized average absorbance from 200 nm to 340 nm ( $n=5$  for each starting material). Circles represent outliers (that lie more than one and a half box lengths above or below from the upper or lower quartile, respectively). Comparison between the starting materials by Kruskal–Wallis test revealed a statistically significant difference ( $p=0.036$ ). Pairwise comparisons of the 6 starting materials were not performed due to the high probability of a type I error when executing 15 tests.

ferences are apparent in both. Additionally, a reproduction of these experiments by another and independent research group would be desirable.

In accordance with our preliminary results,<sup>23</sup> we found a statistically significant difference between the normalized absorbance of solvent control and *A. napellus* 30c globules. The differences in uncorrected absorbance in the present study were less pronounced than in the previous study (data not shown). While the globules in the previous study had been produced for a clinical trial, the ones in the present study were specifically produced for our laboratory study, i.e., control globules and globules impregnated with high dilutions were from the same batch. Thus, if minor impurities existed they would be equally distributed in the globules of all groups and could not account for the differences in absorbance. Since more samples were analyzed, an auto sampler was used in the present study.

In previous studies, we compared UV absorbance of high dilutions of sulfur and quartz to succussed but not diluted water controls. For sulfur, we found statistically significant differences between the high dilutions (6c to 30c combined) and controls on 2 out of 5 measurement days.<sup>12</sup> When each dilution level from 6c to 30c was tested individually against the controls, 16c and 29c had significantly different absorbance even after Bonferroni correction for multiple testing. In the present study, sulfur 200CF differed significantly from the solvent control globules. For quartz, we found no differences between any of the dilution levels and the solvent control. Similarly, in a previous experiment there was no difference between quartz and the respective control.<sup>11</sup> Also in accordance with our previous studies,<sup>10,11</sup> we found no significant differences between UV absorbance of decimal dilutions and controls.

In clinical studies, placebos are used as control interventions and are defined as substances with no pharmacological effect. In

homeopathic in vitro, plant and animal studies several possible controls can be used: the same solvents with which the high dilutions were prepared (e.g., water or water/ethanol),<sup>29–31</sup> succussed but not diluted solvents,<sup>10,30</sup> or diluted and succussed solvents can serve as controls.<sup>30,32,33</sup> Particularly, the succussion may lead to several effects such as ion leaching and gas exchange and in turn altering the properties of the dilutions. Therefore, pure, i.e., unsuccessed solvent is not regarded as adequate when used as single control.<sup>30,34</sup> Diluted and succussed solvents also show effects different from the undiluted solvents.<sup>29,31</sup>

Two different controls were used in this study, native globules and globules impregnated with an ethanol/water mixture that had been succussed (solvent control globules). The normalized average absorbance of these controls differed between the two. Solvent control globules were considered as the more appropriate control and thus used for comparisons with the high dilutions (e.g., in Table 2).

Physico-chemical properties of globules have rarely been investigated. Abel et al. compared  $T_1$  relaxation times of globules impregnated with *Betula alba* 30c and dissolved in water to control globules and found no differences between the two.<sup>35</sup> Sukul et al. prepared potassium bromide pellets from several high dilutions and solvents as replacement for sucrose globules and reported significant differences in Fourier transform infrared spectra between the high dilutions and between the high dilutions and the solvents.<sup>36</sup> Detecting delayed luminescence, Lenger et al. showed differences between homeopathic argentum metallicum globules and controls<sup>37</sup> and found that the stability of ethanolic argentum metallicum declined within a month, while globules were stable over 1 year.<sup>38</sup>

While we found significant differences between potencies of 3 starting materials and controls in this study, 3 other starting materials did not produce such differences. This might not seem

to be in accordance with the hypothesis of a change in the solvent's structure; however, we would not expect the same resulting structures for all starting materials. Therefore, it may well be that some changes are not detectable with our measurement system.

Unexpectedly, we found differences in absorbance between native globules used as controls for the various starting materials, while no difference was detected between solvent control globules (Table 3). This can presently not be explained and may be a result obtained by chance. It may also raise questions about the possible transfer of a therapeutically active ingredient from high dilutions to a control, a phenomenon that has been observed but not yet investigated systematically.

For example, Endler et al.<sup>39</sup> observed an effect of thyroxine 30c on the development of frogs, even when the animals were in the water and had no direct contact to the thyroxine dilution. Variations in controls have been observed before in one of our previous studies with liquid high dilutions.<sup>12</sup> However, the variations in the present and previous studies occurred irregularly and did not indicate a systematic influence of high dilutions on controls.

As described above, several modes of action were proposed that lead to a change in water structure during potentization, but it is yet unclear how a changed structure may be preserved on globules. As an explanation the importance of lattice defects in lactose monohydrate or small pores on sucrose globules to transfer therapeutically active ingredients from high dilutions has been pointed out.<sup>40</sup> It has also been suggested that in the solid phase, nanostructures may retain their properties without dissipating energy from the environment and could return to their preceding state when water became available,<sup>41</sup> but precise models of this process are lacking. Therefore, it is important to gain insights in physical properties of globules as these may facilitate to developing models to better understanding their effects and modes of action. Finally, one of the next achievements will be to transfer findings from such physico-chemical measurements on effects observed in plants, animals and humans.

## 5. Conclusions

Globules prepared from high dilutions of *A. napellus*, sulfur and *A. mellifica* showed significantly different UV absorbance compared to solvent control globules when dissolved in water. This study has expanded our findings from aqueous high dilutions to globules, and it suggests that characteristics of aqueous high dilutions may be preserved and detectable in dissolved globules.

## Conflict of interest

None.

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